

# SARS-COV-2, INFLUENZA A AND INFLUENZA B MULTIPLEX qPCR TEST KIT USER GUIDE

CAT NO.: YSL-qPX-EC-COV.FLUAB-100

100 reactions

with Endogenous Control and Lyophilised MasterMix

## **VERSION 3.5**

For research use only



#### YouSeg Ltd

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# INTENDED USE

This product is a qPCR test kit for detection of COVID19 (Covid19) RNA, Influenza A (FluA) RNA and Influenza B (FluB) RNA in good quality nucleic acid samples from a variety of sources. It is designed to be used by trained users in a suitable molecular biology laboratory environment.

## KIT CONTENTS

	Cap Colour	Volume
Covid19 specific primer/probe (VIC/HEX Probe) FluA specific primer/probe (FAM Probe) FluB specific primer/probe (ROX Probe) Endogenous control primer/probe (CY5 Probe)		11Ο μΙ
Lyophilised Tetra OneStep 2X qRT-PCR MasterMix		1.1 ml*
COV.FLUAB positive control template		500 μΙ*
MasterMix resuspension buffer		1.5 ml
DNase/RNase free water		1.5 ml
Template resuspension buffer		1.5 ml

<sup>\*</sup> Supplied lyophilised and requires resuspension before use, see resuspension step below for instructions

# **RESUSPENSION**

Resuspend the designated kit contents with the correct reagents as per the table below. Spin or gently tap the vials to ensure all the contents is at the bottom before opening.

After adding the resuspension reagent, pulse vortex the vials to ensure it is mixed well.

\\ r \ \ r \///////	Reagent	Volume
Lyophilised Tetra OneStep 2X qRT-PCR MasterMix	MasterMix resus. buffer	1.1 ml
COV.FLUAB positive control template	Template resus. buffer	500 μΙ



## MATERIALS REQUIRED BUT NOT PROVIDED

RNA Extraction kit - This qPCR test kit will work well with high quality RNA derived from any extraction kit with minimal PCR inhibitors present.

qPCR instrument with minimum 4 colour detection (VIC/HEX, FAM, ROX and CY5).

Pipettes, micro centrifuge tubes and general laboratory equipment.

## KIT SPECIFICITY

The YouSeq qPCR test kit for detection of SARS-CoV-2, Influenza A and Influenza B (COV.FLUAB) is designed to have the broadest detection profile possible and detect all clinically relevant strains. The primers and probes have very high (>95%) homology with all reference data within the NCBI database.

The target genes for COVID19, Influenza A and Influenza B (N & RdRp genes, M1 gene and NEP/NS1 gene respectively) have been demonstrated to have unique sequences within these species making them ideal targets for highly specific detection of these pathogens.

If you require more specific data about the detection profile of the kit, please do not hesitate to contact our bioinformatics team: support@youseq.com

# qRT-PCR BENCH SIDE PROTOCOL

Clean and decontaminate all work surfaces, pipettes and other equipment prior to use to remove potentially contaminating nucleic acids.

#### REACTION SET UP •

Combine the following reagents to create a final test reaction:

Component	Volume
Tetra OneStep 2X qRT-PCR MasterMix	10 μΙ
COV.FLUAB specific primer/probe	1 μΙ
Extracted Sample RNA	9 μΙ
Final Volume	20 μΙ

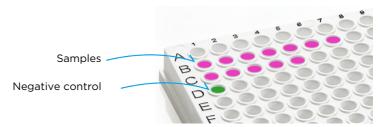
Please note: Work swiftly and on ice. YouSeq Tetra OneStep qRT-PCR MasterMix contains a powerful reverse transcriptase enzyme to deliver maximally efficient conversion of viral RNA to cDNA. This enzyme is active at room temperature. If left at room temperature in the presence of primers/probes the reverse transcriptase can react with the primers and probe to create artefacts that reduce assay sensitivity. Therefore, it is critical to store your primer/probe and MasterMix reaction mix on ice and for periods of no more than 30 minutes.

For the same reason, set up your qPCR reaction plate on ice and proceed to amplification within 30 minutes. Do not delay.



#### **NEGATIVE CONTROL**

For a negative control reaction, repeat the reaction set up above replacing the sample RNA with DNase/RNase free water.



Please note: Make sure to seal the sample and negative control wells before proceeding to the positive control step.

### POSITIVE CONTROL

For a positive control reaction, repeat the reaction set up above replacing the sample RNA with 9  $\mu$ l of the positive control template supplied with the kit.

# qPCR AMPLIFICATION PROTOCOL

Run the following PCR protocol:

Please note: If using a qPCR machine that uses ROX as a passive reference, then the passive reference must be turned off or set to "none" indicating no passive reference.

, /	Temperature	Time	
RT Step	55°C	10 minutes	
Hot Start	95°C	3 minutes	
45 cycles	95°C	15 seconds	
43 Cycles	60°C*	60 seconds	

<sup>\*</sup>Make sure to collect fluorogenic data through VIC/HEX, FAM, ROX and CY5 channels during this step



## INTERPRETATION OF RESULTS

When analysing Sample Cq values, YouSeq recommends checking the threshold within the run file before interpreting the data. We would suggest setting the threshold to 10% of the relevant positive control End Point Fluorescence (EPF).

#### **Positive control**

Firstly, check the positive control performance. The positive control should amplify in a Cq range of approximately 18.5 +/-2. If the Cq range is not achieved, this would be a failed test and should be repeated.

Please note: The positive control in the kit is a representative sequence associated to the designs target region and does not contain the organisms entire genome.

The positive control within the kit does not include the endogenous control sequence. Therefore, the positive control should not be expected to amplify in the endogenous control channel.

#### **Negative control**

In ideal circumstances, the negative control should deliver a flat line – negative result. However, it is not uncommon for background laboratory contamination to cause a very late signal. If this signal is  $\geq 5$  Cq values later than your sample signal then it can be considered negative and the result is viable.

If the negative control is <5 Cq later than the signal sample, then the result is inconclusive and the test should be repeated after potential sources of contamination have been removed.

The test is valid if the following conditions are met:

	VIC/HEX Channel	FAM Channel	ROX Channel
Positive Control	+	+	+
Negative Control	-	-	<del>-</del>

#### **Positive samples**

Samples that are positive for Covid19, FluA or FluB will deliver a defined "sigmoidal" amplification plot.

#### **Endogenous control**

If your sample delivers a strong positive result then the endogenous control is not required for data interpretation and can be ignored.

If your samples deliver a negative result, then the endogenous control is useful to interpret the result. The Cq value from the endogenous control will vary according to the amount of RNA in your sample. A late signal (Cq>28) indicates that only a small amount of host derived RNA was present in your sample. You may wish to repeat sample collection and then repeat the test in order to confirm the negative result.



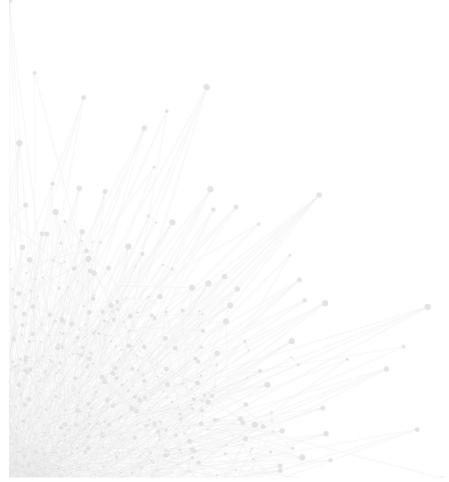
# INTERPRETATION OF RESULTS CONTINUED

#### Results interpretation at a glance:

		qPCR Signal			
Covid19 Sample	+	-	-	-	-
FluA Sample	-	+	-	-	-
FluB Sample	-	-	+	-	-
Endogenous Control	+/-	+/-	+/-	+	-
Result	Positive result Covid19 specific RNA detected	Positive result FluA specific RNA detected	Positive result FluB specific RNA detected	Negative Result	Failed test. Insufficient RNA

#### **Coinfection:**

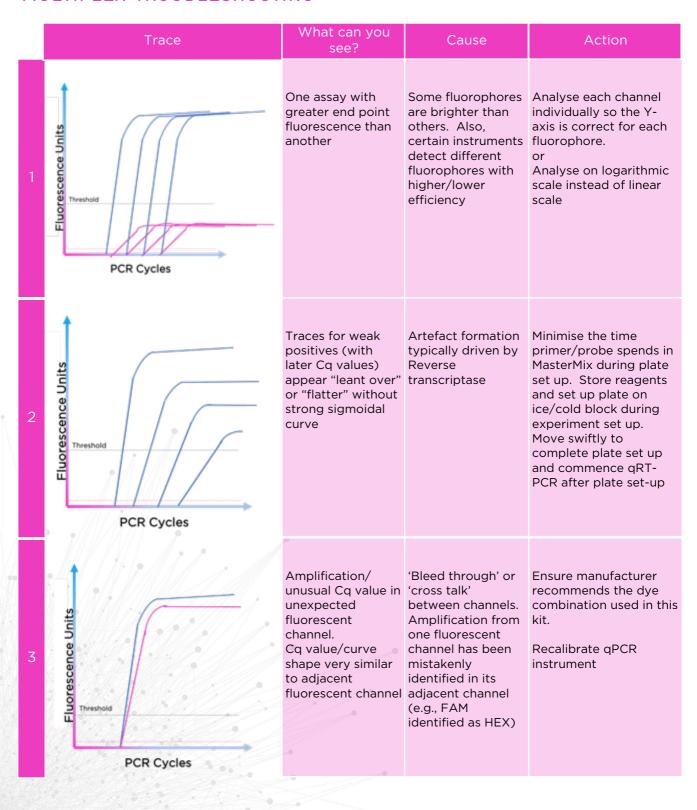
On the rare occasion that a sample contains more than one target pathogen, positive signals in multiple channels will be observed.



Page 6



# MULTIPLEX TROUBLESHOOTING





# PRODUCT SPECIFICATIONS

#### Storing your kit

Store at -20°C from arrival. The qPCR kits shelf life is outlined as an expiry date on the pouch label.

#### **Use good quality RNA**

Poor quality input nucleic acid is the biggest cause of test failure. The kit will work well with any source of good quality RNA. Good quality is defined as RNA with high integrity (not degraded) and with low levels of inhibitors present.

#### **Regulatory status**

This product has been developed for Research Use Only and is not intended for diagnostic use. It should not be used for diagnosis of disease unless specifically approved by the regulatory authorities in the country of use.

#### **Quality Control**

In accordance with the YouSeq Ltd ISO EN 13485-certified Quality Management System, each lot of SARS-CoV-2, Influenza A and Influenza B Multiplex qPCR kit is tested against predetermined specifications to ensure consistent product quality. Design of the kit met our robust bioinformatic analysis requirements resulting in a clinically relevant detection profile based on available sequence information. The kit is periodically checked against newly available sequence information to remain clinically relevant.

#### **Technical Assistance**

For customer support, please contact:

e-mail: support@youseq.com phone: +44 (0)333 577 6697

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